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(54) Title: IkB KINASES

(57) Abstract

The invention provides methods and compositions relating to InB regulating proteins, known as T2K proteins, and related nucleic acids. The proteins may be produced recombinantly from transformed host cells from the disclosed T2K encoding nucleic acid or purified from human cells. The invention provides specific hybridization probes and primers capable of specifically hybridizing with the disclosed T2K gene, T2K—specific binding agents such as specific antibodies, and methods of making and using the subject compositions in diagnosis, therapy and in the biopharmaceutical industry.

IKB Kinases

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INTRODUCTION

Field of the Invention

The field of this invention is a family of kinases which regulate signal transduction.

Background

Inflammatory cytokines IL-1 and TNF exert diverse biological activities by altering gene expression in the cells, a function mediated mostly by transcription factor NF-κB. In unstimulated cells, NF-κB proteins form a complex with inhibitory molecules, the IκB proteins, and are rendered inactive in the cytoplasm. In response to cytokines and other stimuli, the IκB proteins are phosphorylated on specific serine residues. In particular, phosphorylation of two serine residues as part of the consensus sequence DSGL/IXSM/L (e.g. ser 32 and 36 in IκBα, ser 19 and 23 in IκBβ, and ser 157 and 161, or 18 and 22, depending on the usage of methionines, in IκBε, respectively) which mark the proteins for ubiquitination and proteosome-mediated degradation, releasing NF-κB to enter the nucleus to activate the genes that encode proteins participating in inflammatory and immune responses. Henceforth, the term IκB serine 36 is used herein to refer generically to the second serine residue of the foregoing consensus sequence, e.g. that corresponding to serine 36 in IκBα, ser 23 in IκBβ, and ser 161 or 22 in IκBε.

Delineating TNF and IL-1 signaling pathways for NF-kB activation has implicated the TRAF molecules as converging point for different cytokines, with TRAF2 being involved in TNF- and TRAF6 in IL-1-induced NF-kB activation. We disclose herein a family of IkB kinases including a TRAF2-associated kinase activity (designated T2K) and the translation product of the KIAA0151 gene product that phosphorylates the IkB molecules on the specific regulatory serine residues. We also disclose the purification of a native protein responsible for such kinase activity, the sequencing of T2K peptides derived, and the cloning of native T2K cDNA.

Relevant Literature

Nagase et al. (1995) DNA Res. 2(4),167-174 report conceptual coding sequences from a number of unidentified human genes including KIAA0151. Song et al., US Patent Application Serial No. 08/677,862 discloses a TRAF2-associated kinase.

SUMMARY OF THE INVENTION

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The invention provides methods and compositions relating to natural isolated regulatory proteins called T2K proteins, related nucleic acids, and protein domains thereof having T2K-specific activity. The proteins may be produced recombinantly from transformed host cells from the subject T2K encoding nucleic acids or purified from mammalian cells. The invention provides isolated T2K hybridization probes and primers capable of specifically hybridizing with the disclosed T2K gene, T2K-specific binding agents such as specific antibodies, and methods of making and using the subject compositions in diagnosis (e.g. genetic hybridization screens for T2K transcripts), therapy (e.g. gene therapy to modulate T2K gene expression) and in the biopharmaceutical industry (e.g. as immunogens, reagents for isolating other transcriptional regulators, reagents for screening chemical libraries for lead pharmacological agents, etc.).

DETAILED DESCRIPTION OF THE INVENTION

The nucleotide sequences of a natural cDNA encoding a human T2K protein is shown as SEQ ID NO:1, and the full conceptual translate is shown as SEQ ID NO:2. The T2K proteins of the invention include incomplete translates of SEQ ID NO:1 and deletion mutants of SEQ ID NO:2, which translates and deletion mutants have T2K-specific amino acid sequence and assay-discernable T2K-specific binding specificity or function. Such active T2K deletion mutants, T2K peptides or protein domains comprise a sequence of at least about 6, preferably at least about 8, more preferably at least about 10 consecutive residues of SEQ ID NO:2 which distinguishes both the KIAA0151 gene product and the translation product of SEQ ID NO:1, bases 1756-2095. For examples, T2K protein domains identified below are shown to provide protein-binding domains which are identified in and find use, *inter alia*, in solid-phase binding and kinase assays as described below.

T2K-specific activity or function may be determined by convenient *in vitro*, cell-based, or *in vivo* assays: e.g. *in vitro* binding assays, cell culture assays, in animals (e.g. gene therapy, transgenics, etc.), etc. Binding assays encompass any assay where the molecular

interaction of an T2K protein with a binding target is evaluated. The binding target may be a natural intracellular binding target (including substrates, agonists and antagonists) such as an IkB or TRAF2, or other regulator that directly modulates T2K activity or its localization; or non-natural binding target such a specific immune protein such as an antibody, or an T2K specific agent such as those identified in screening assays such as described below. T2K-binding specificity may assayed by binding equilibrium constants (usually at least about 10⁷ M⁻¹, preferably at least about 10⁸ M⁻¹, more preferably at least about 10⁹ M⁻¹), by the ability of the subject protein to function as negative mutants in T2K-expressing cells, to elicit T2K specific antibody in a heterologous host (e.g a rodent or rabbit), etc.; or, in a preferred embodiment, by kinase activity.

The claimed T2K proteins are isolated or pure: an "isolated" protein is unaccompanied by at least some of the material with which it is associated in its natural state, preferably constituting at least about 0.5%, and more preferably at least about 5% by weight of the total protein in a given sample and a pure protein constitutes at least about 90%, and preferably at least about 99% by weight of the total protein in a given sample. The T2K proteins and protein domains may be synthesized, produced by recombinant technology, or purified from mammalian, preferably human cells. A wide variety of molecular and biochemical methods are available for biochemical synthesis, molecular expression and purification of the subject compositions, see e.g. Molecular Cloning, A Laboratory Manual (Sambrook, et al. Cold Spring Harbor Laboratory), Current Protocols in Molecular Biology (Eds. Ausubel, et al., Greene Publ. Assoc., Wiley-Interscience, NY) or that are otherwise known in the art.

The invention provides natural and non-natural T2K-specific binding agents, methods of identifying and making such agents, and their use in diagnosis, therapy and pharmaceutical development. For example, T2K-specific agents are useful in a variety of diagnostic and therapeutic applications. Novel T2K-specific binding agents include T2K-specific receptors, such as somatically recombined protein receptors like specific antibodies or T-cell antigen receptors (see, e.g Harlow and Lane (1988) Antibodies, A Laboratory Manual, Cold Spring Harbor Laboratory) and other natural intracellular binding agents identified with assays such as one-, two- and three-hybrid screens, non-natural intracellular binding agents identified in screens of chemical libraries such as described below, etc. For diagnostic uses, the binding agents are frequently labeled, such as with fluorescent, radioactive, chemiluminescent, or other easily detectable molecules, either conjugated directly to the binding agent or

conjugated to a probe specific for the binding agent. Agents of particular interest modulate T2K function, e.g. T2K kinase activity; for example, isolated cells, whole tissues, or individuals may be treated with a T2K binding agent to activate, inhibit, or alter T2K-kinase dependent processes such as NfkB activation.

The amino acid sequences of the disclosed T2K proteins are used to back-translate T2K protein-encoding nucleic acids optimized for selected expression systems (Holler et al. (1993) Gene 136, 323-328; Martin et al. (1995) Gene 154, 150-166) or used to generate degenerate oligonucleotide primers and probes for use in the isolation of natural T2K-encoding nucleic acid sequences ("GCG" software, Genetics Computer Group, Inc, Madison WI). T2K-encoding nucleic acids used in T2K-expression vectors and incorporated into recombinant host cells, e.g. for expression and screening, transgenic animals, e.g. for functional studies such as the efficacy of candidate drugs for disease associated with T2K-modulated cell function, etc.

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The invention also provides nucleic acid hybridization probes and replication / amplification primers having a hitherto novel T2K cDNA specific sequence contained in SEQ ID NO:1 (including its complement and analogs and complements thereof having the corresponding sequence, e.g. in RNA) and sufficient to effect specific hybridization thereto (i.e. specifically hybridize with SEQ ID NO:1 in the presence of the KIAA0151 gene and nucleic acids consisting of SEO ID NO:1, bases 1756-2095). Such primers or probes are at least 12, preferably at least 24, more preferably at least 36 and most preferably at least 96 bases in length. Demonstrating specific hybridization generally requires stringent conditions, for example, hybridizing in a buffer comprising 30% formamide in 5 x SSPE (0.18 M NaCl, 0.01 M NaPO₄, pH7.7, 0.001 M EDTA) buffer at a temperature of 42°C and remaining bound when subject to washing at 42°C with 0.2 x SSPE; preferably hybridizing in a buffer comprising 50% formamide in 5 x SSPE buffer at a temperature of 42°C and remaining bound when subject to washing at 42°C with 0.2 x SSPE buffer at 42°C. T2K cDNA homologs can also be distinguished from other protein using alignment algorithms, such as BLASTX (Altschul et al. (1990) Basic Local Alignment Search Tool, J Mol Biol 215, 403-410).

The subject nucleic acids are of synthetic/non-natural sequences and/or are isolated, i.e. unaccompanied by at least some of the material with which it is associated in its natural state, preferably constituting at least about 0.5%, preferably at least about 5% by weight of

total nucleic acid present in a given fraction, and usually recombinant, meaning they comprise a non-natural sequence or a natural sequence joined to nucleotide(s) other than that which it is joined to on a natural chromosome. Nucleic acids comprising the nucleotide sequence of SEQ ID NO:1 or fragments thereof, contain such sequence or fragment at a terminus, immediately flanked by a sequence other than that which it is joined to on a natural chromosome, or flanked by a native flanking region fewer than 10 kb, preferably fewer than 2 kb, which is at a terminus or is immediately flanked by a sequence other than that which it is joined to on a natural chromosome. While the nucleic acids are usually RNA or DNA, it is often advantageous to use nucleic acids comprising other bases or nucleotide analogs to provide modified stability, etc.

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The subject nucleic acids find a wide variety of applications including use as translatable transcripts, hybridization probes, PCR primers, diagnostic nucleic acids, etc.; use in detecting the presence of T2K genes and gene transcripts and in detecting or amplifying nucleic acids encoding additional T2K homologs and structural analogs. In diagnosis, T2K hybridization probes find use in identifying wild-type and mutant T2K alleles in clinical and laboratory samples. Mutant alleles are used to generate allele-specific oligonucleotide (ASO) probes for high-throughput clinical diagnoses. In therapy, therapeutic T2K nucleic acids are used to modulate cellular expression or intracellular concentration or availability of active T2K. For example, T2K nucleic acids are also used to modulate cellular expression or intracellular concentration or availability of active T2K protein. T2K inhibitory nucleic acids are typically antisense: single-stranded sequences comprising complements of the disclosed natural T2K coding sequences. Antisense modulation of the expression of a given T2K protein may employ antisense nucleic acids operably linked to gene regulatory sequences. Cell are transfected with a vector comprising a T2K sequence with a promoter sequence oriented such that transcription of the gene yields an antisense transcript capable of binding to endogenous T2K encoding mRNA. Transcription of the antisense nucleic acid may be constitutive or inducible and the vector may provide for stable extrachromosomal maintenance or integration. Alternatively, single-stranded antisense nucleic acids that bind to genomic DNA or mRNA encoding a given T2K protein may be administered to the target cell, in or temporarily isolated from a host, at a concentration that results in a substantial reduction in expression of the targeted protein. An enhancement in T2K expression is effected by introducing into the targeted cell type T2K nucleic acids which increase the

functional expression of the corresponding gene products. Such nucleic acids may be T2K expression vectors, vectors which upregulate the functional expression of an endogenous allele, or replacement vectors for targeted correction of mutant alleles. Techniques for introducing the nucleic acids into viable cells are known in the art and include retroviral-based transfection, viral coat protein-liposome mediated transfection, etc.

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The invention provides efficient methods of identifying agents, compounds or lead compounds for agents active at the level of an IkB serine 36 specific kinase protein modulatable cellular function. Generally, these screening methods involve assaying for compounds which modulate IkB serine 36 specific kinase protein interaction with a natural IkB serine 36 specific kinase protein binding target. A wide variety of assays for binding agents are provided including labeled *in vitro* protein-protein binding assays, immunoassays, cell based assays, etc. The methods are amenable to automated, cost-effective high throughput screening of chemical libraries for lead compounds. Such libraries encompass candidate agents of encompass numerous chemical classes, though typically they are organic compounds; preferably small organic compounds and are obtained from a wide variety of sources including libraries of synthetic or natural compounds. Identified agents find use in the pharmaceutical industries for animal and human trials; for example, the agents may be derivatized and rescreened in *in vitro* and *in vivo* assays to optimize activity and minimize toxicity for pharmaceutical development.

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In vitro binding assays employ a mixture of components including a IkB serine 36 specific kinase protein such as a T2K protein, which may be part of a fusion product with another peptide or polypeptide, e.g. a tag for detection or anchoring, etc. The assay mixtures comprise a natural intracellular binding target of the kinase protein. While native binding targets may be used, it is frequently preferred to use portions (e.g. peptides) thereof so long as the portion provides binding affinity and avidity to the subject kinase protein conveniently measurable in the assay. In a particular embodiment, the binding target is a substrate comprising IkB serine 36. Such substrates comprise a IkB α , β or ϵ peptide including the serine 36 residue and at least 5, preferably at least 10, and more preferably at least 20 naturally occuring immediately flanking residues on each side (i.e. residues 26-46, 22-42, or 12-32 or 151-171 for IkB α , β or ϵ -derived substrates, respectively).

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The assay mixture also comprises a candidate pharmacological agent and typically, a variety of other reagents such as salts, buffers, neutral proteins, e.g. albumin, detergents,

protease inhibitors, nuclease inhibitors, antimicrobial agents, etc. The mixture components can be added in any order that provides for the requisite bindings and incubations may be performed at any temperature which facilitates optimal binding. The mixture is then incubated under conditions whereby, but for the presence of the candidate pharmacological agent, the kinase protein specifically binds the cellular binding target, portion or analog with a reference binding affinity. Incubation periods are likewise selected for optimal binding but also minimized to facilitate rapid, high-throughput screening.

After incubation, the agent-biased binding between the kinase protein and one or more binding targets is detected by any convenient way. First, a separation step is generally used to separate bound from unbound components. Separation may be effected by precipitation (e.g. TCA precipitation, immunoprecipitation, etc.), immobilization (e.g on a solid substrate), etc., followed by washing by, for examples, membrane filtration, gel chromatography (e.g. gel filtration, affinity, etc.). One of the components usually comprises or is coupled to a label. The label may provide for direct detection as radioactivity, luminescence, optical or electron density, etc. or indirect detection such as an epitope tag, an enzyme, etc. A variety of methods may be used to detect the label depending on the nature of the label and other assay components, e.g. through optical or electron density, radiative emissions, nonradiative energy transfers, etc. or indirectly detected with antibody conjugates, etc. A difference in the binding affinity of the kinase protein to the target in the absence of the agent as compared with the binding affinity in the presence of the agent indicates that the agent modulates the binding of the kinase protein to the binding target. Analogously, in the cell-based transcription assay also described below, a difference in the kinase protein transcriptional induction in the presence and absence of an agent indicates the agent modulates kinase-modulated transcription. A difference, as used herein, is statistically significant and preferably represents at least a 50%, more preferably at least a 90% difference.

The following experimental section and examples are offered by way of illustration and not by way of limitation.

EXPERIMENTAL

Identification of T2K:

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293 cells were stably transfected with DNA plasmids that direct the expression of the human TRAF2 protein with an N-terminal Flag-epitope tag. Cells grown in suspension culture

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were pelleted in 500 ml bottles in a Sorvall GS-3 rotor spun at 2000 RPM for 5 minutes and were lysed in 5 pelleted-cell-volumes of "lysis buffer" containing 50 mM Hepes, pH 7.9, 250 mM NaCl, 5 mM dithiothreitol (DTT), 1 mM EDTA, 20 mM β glycerophosphate, 5 mM p-nitrophenyl phosphate, 1 mM Na orthovanadate, 1 mM benzamidine, 0.4 mM phenylmethylsulfonyl fluoride (PMSF), 1 mM Na metabisulfite, 0.1% NP-40 and 10% (v/v) glycerol. After incubation on ice for 30 minutes with occasional rocking, cell lysate was centrifuged in a 50 ml conical tube in a Sorvall H6000A rotor at 4000 RPM for 10 minutes. Supernatants were collected and centrifuged in a Beckman 45 TI rotor at 40,000 RPM for 2 hours. The TRAF complex was immunoprecipitated using anti-flag monoclonal antibodies cross-linked to sepharose (VWR) (1.5 ml sepharose beads for 200 mls of extracts). The immunoprecipitates were washed 4 times with cell lysis buffer, twice with lysis buffer containing 1 molar NaCl, then twice with lysis buffer. At this stage, the immunocomplex can efficiently phosphorylate wild type $I\kappa B\alpha$ and β but not the mutants with the two serines substituted with alanines. The sepharose beads containing TRAF2 complex were then incubated at 30 °C for 1 hour in 4.5 mls of kinase buffer containing 20 mM Tris-HCl, pH 7.6, 20 mM MgCl₂, 20 mM β glycerophosphate, 20 mM p-nitrophenyl phosphate, 1 mM EDTA, 1 mM Na orthovanadate, 1 mM benzamidine, 0.4 mM PMSF, 1 mM Na metabisulfite, 1 mM ATP, and 20 mM creatinephosphate. After the in vitro kinase reaction, significant amounts of the IkB kinase activity were found in the soluble fraction which was loaded on an 1 ml heparin agarose column and eluted with a NaCl gradient. The IkB kinase activity was recovered in the flow though fraction which was concentrated with a centricon (Amicon) to 50 ul. The material was fractionated on a superdex 200 gel filtration column driven by the Smart system (Pharmacia) and the eluate was collected in 50 ul fractions. The kinase activity was recovered in the fractions that correlated with molecule size marker of 670 kD. These fractions were pooled and further separated on a Mono Q column by a NaCl linear gradient. The kinase activity was found in 0.3 to 0.4 M NaCl eluate. Silver staining of the column fractions separated on SDS gels revealed an 85 to 90 kD polypeptide that correlated with the kinase activity in both superdex 200 and Mono Q fractionation. After SDS gel separation, this polypeptide was subjected to micropeptide sequencing. One peptide sequence obtained matched a partial cDNA sequence in the Merck-Washington University Est database. A cDNA clone that contains open reading frame for 729 amino acids was isolated from a lambda phage cDNA library generated from HeLa cells. Sequence analysis revealed a protein kinase domain in the N-terminal portion of the predicted

protein (T2K). Searching protein sequence database with the kinase domain of T2K identified a protein (KIAA0151) highly homologous to T2K, specially in the protein kinase domain (75% identity). KIAA0151 is a kinase with undefined function and was reported by Nagase T. *et al.* as a novel cDNA sequence isolated from human KG-1 cells (DNA Res. 2 (4), 167-174 (1995).

Substrate specificity analysis revealed that both T2K and KIAA0151 specifically phosphorylate IkB serine 36 and associate with TRAF2. Furthermore, deletion mutant analysis reveals that residues 10-250 define kinase domains and residues 251-729 and 251-716, for T2K and KIAA0151 respectively, define regulatory domains active as a negative mutants for IkB kinase activity. Recombinant T2K kinase is prepared by over-expressing GST fusion proteins in E. coli and baculavirus expression systems.

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EXAMPLES

1. Protocol for at T2K - $I\kappa B\alpha$ phosphorylation assay.

A. Reagents:

- Neutralite Avidin: 20 μg/ml in PBS.
- kinase: 10^{-8} 10^{-5} M kinase (SEQ ID NO:2) at 20 μ g/ml in PBS.
- <u>substrate</u>: 10^{-7} 10^{-4} M biotinylated substrate (21 residue peptide consisting of residues 26-46 of human $I\kappa B\alpha$) at 40 μ g/ml in PBS.
 - Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.
- Assay Buffer: 100 mM KCl, 10 mM MgCl₂, 1 mM MnCl₂, 20 mM HEPES pH 7.4, 0.25 mM EDTA, 1% glycerol, 0.5% NP-40, 50 mM BME, 1 mg/ml BSA, cocktail of protease inhibitors.
- -[32 P] γ -ATP 10x stock: 2 x 10⁵M cold ATP with 100 μ Ci [32 P] γ -ATP. Place in the 4°C microfridge during screening.
- Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506), 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVo₃ (Sigma # S-6508) in 10 ml of PBS.
- B. Preparation of assay plates:
 - Coat with 120 µl of stock N Avidin per well overnight at 4°C.
 - Wash 2 times with 200 µl PBS.
 - Block with 150 µl of blocking buffer.
 - Wash 2 times with 200 µl PBS.

C. Assay:

- Add 40 µl assay buffer/well.
- Add 40 µl biotinylated substrate (2-200 pmoles/40 ul in assay buffer)
- Add 40 µl kinase (0.1-10 pmoles/40 ul in assay buffer)
- Add 10 µl compound or extract.
- Add 10 μl [³²P]γ-ATP 10x stock.
 - Shake at 25°C for 15 minutes.
 - Incubate additional 45 minutes at 25°C.
 - Stop the reaction by washing 4 times with 200 μ l PBS.
 - Add 150 µl scintillation cocktail.
- 10 Count in Topcount.
 - D. Controls for all assays (located on each plate):
 - a. Non-specific binding
 - b. cold ATP at 80% inhibition.
- 15 2. Protocol for at KIAA0151 IκBβ phosphorylation assay.
 - A. Reagents:
 - Neutralite Avidin: 20 µg/ml in PBS.
 - kinase: 10^{-8} 10^{-5} M truncated KIAA0151 kinase (residues 4-714) at 20 μ g/ml in PBS.
 - <u>substrate</u>: 10⁻⁷ 10⁻⁴ M biotinylated substrate (21 residue peptide consisting of residues
- 20 22-42 of human $I\kappa B\beta$) at 40 μ g/ml in PBS.
 - Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.
 - Assay Buffer: 100 mM KCl, 10 mM MgCl₂, 1 mM MnCl₂, 20 mM HEPES pH 7.4, 0.25 mM EDTA, 1% glycerol, 0.5% NP-40, 50 mM BME, 1 mg/ml BSA, cocktail of protease inhibitors.
- 25 -[32P]γ-ATP 10x stock: 2 x 10-5 M cold ATP with 100 μCi [32P]γ-ATP. Place in the 4°C microfridge during screening.
 - Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506), 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVo₃ (Sigma # S-6508) in 10 ml of PBS.
- 30 B. Preparation of assay plates:
 - Coat with 120 µl of stock N Avidin per well overnight at 4°C.

- Wash 2 times with 200 µl PBS.
- Block with 150 µl of blocking buffer.
- Wash 2 times with 200 µl PBS.

C. Assay:

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- Add 40 µl assay buffer/well.
- Add 40 µl biotinylated substrate (2-200 pmoles/40 ul in assay buffer)
- Add 40 µl kinase (0.1-10 pmoles/40 ul in assay buffer)
- Add 10 µl compound or extract.
- Add 10 μ l [³²P] γ -ATP 10x stock.
- Shake at 25°C for 15 minutes.
- Incubate additional 45 minutes at 25°C.
 - Stop the reaction by washing 4 times with 200 µl PBS.
 - Add 150 µl scintillation cocktail.
 - Count in Topcount.
 - D. Controls for all assays (located on each plate):
 - a. Non-specific binding
 - b. cold ATP at 80% inhibition.
 - 3. Protocol for high throughput T2K-TRAF2 heterodimer formation assay.
 - A. Reagents:
- 20 <u>Neutralite Avidin</u>: 20 μg/ml in PBS.
 - Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.
 - Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 1 mM MgCl₂, 1% glycerol, 0.5% NP-40, 50 mM β-mercaptoethanol, 1 mg/ml BSA, cocktail of protease inhibitors.
 - ³³P T2K protein 10x stock: 10⁻⁸ 10⁻⁶ M "cold" T2K supplemented with 200,000-250,000 cpm of labeled T2K (Beckman counter). Place in the 4°C microfridge during screening.
 - Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506), 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVo₃ (Sigma # S-6508) in 10 ml of PBS.
 - -TRAF2: 10⁻⁷ 10⁻⁵ M biotinylated TRAF2 in PBS.
- 30 B. Preparation of assay plates:
 - Coat with 120 µl of stock N-Avidin per well overnight at 4°C.

- Wash 2 times with 200 µl PBS.
- Block with 150 µl of blocking buffer.
- Wash 2 times with 200 µl PBS.

C. Assay:

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- Add 40 µl assay buffer/well.
- Add 10 µl compound or extract.
 - Add 10 μ l ³³P-T2K (20-25,000 cpm/0.1-10 pmoles/well =10⁻⁹- 10⁻⁷ M final conc).
 - Shake at 25°C for 15 minutes.
 - Incubate additional 45 minutes at 25°C.
 - Add 40 µl biotinylated TRAF2 (0.1-10 pmoles/40 ul in assay buffer)
- Incubate 1 hour at room temperature.
 - Stop the reaction by washing 4 times with 200 μM PBS.
 - Add 150 µM scintillation cocktail.
 - Count in Topcount.
 - D. Controls for all assays (located on each plate):
 - a. Non-specific binding
 - b. Soluble (non-biotinylated TRAF2) at 80% inhibition.

All publications and patent applications cited in this specification are herein incorporated by reference as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference. Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be readily apparent to those of ordinary skill in the art in light of the teachings of this invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims.

(1) GENERAL INFORMATION:

(I) APPLICANT: Cao, Zhaodan

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- (ii) TITLE OF INVENTION: TRAF2-Associated Kinase
- (iii) NUMBER OF SEQUENCES: 2
- 10 (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: SCIENCE & TECHNOLOGY LAW GROUP
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 - (D) STATE: CALIFORNIA
- 15 (E) COUNTRY: USA
 - (F) ZIP: 94104
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
- 20 (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
 - (vi) CURRENT APPLICATION DATA:
- 25 (A) APPLICATION NUMBER: US
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
 - (viii) ATTORNEY/AGENT INFORMATION:
- 30 (A) NAME: OSMAN, RICHARD A
 - (B) REGISTRATION NUMBER: 36,627
 - (C) REFERENCE/DOCKET NUMBER: T97-002
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (415) 343-4341
 - (B) TELEFAX: (415) 343-4342
 - (2) INFORMATION FOR SEQ ID NO:1:
- 40 (I) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2994 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: CDS

5 (B) LOCATION: 73..2259

	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:									
10	GCGGGAGCCC GCCGGCGGTG GCGCGGCGGA GACCCGGCTG GTATAACAAG AGGATTGCCT	60								
10	GATCCAGCCA AG ATG CAG AGC ACT TCT AAT CAT CTG TGG CTT TTA TCT Met Gln Ser Thr Ser Asn His Leu Trp Leu Leu Ser 1 5 10									
15	GAT ATT TTA GGC CAA GGA GCT ACT GCA AAT GTC TTT CGT GGA AGA CAT Asp Ile Leu Gly Gln Gly Ala Thr Ala Asn Val Phe Arg Gly Arg His	156								
	15 20 25									
	AAG AAA ACT GGT GAT TTA TTT GCT ATC AAA GTA TTT AAT AAC ATA AGC	204								
20	Lys Lys Thr Gly Asp Leu Phe Ala Ile Lys Val Phe Asn Asn Ile Ser									
	30 35 40									
	TTC CTT CGT CCA GTG GAT GTT CAA ATG AGA GAA TTT GAA GTG TTG AAA	252								
25	Phe Leu Arg Pro Val Asp Val Gln Met Arg Glu Phe Glu Val Leu Lys									
25	45 . 50 . 55 . 60									
	AAA CTC AAT CAC AAA AAT ATT GTC AAA TTA TTT GCT ATT GAA GAG GAG	300								
	Lys Leu Asn His Lys Asn Ile Val Lys Leu Phe Ala Ile Glu Glu									
30	65 70 75									
50	ACA ACA ACA AGA CAT AAA GTA CTT ATT ATG GAA TTT TGT CCA TGT GGG	348								
	Thr Thr Thr Arg His Lys Val Leu Ile Met Glu Phe Cys Pro Cys Gly									
	80 85 90									
35	AGT TTA TAC ACT GTT TTA GAA GAA CCT TCT AAT GCC TAT GGA CTA CCA	396								
	Ser Leu Tyr Thr Val Leu Glu Glu Pro Ser Asn Ala Tyr Gly Leu Pro									
	95 100 105									
	GAA TCT GAA TTC TTA ATT GTT TTG CGA GAT GTG GTG GGT GGA ATG AAT	444								
40	Glu Ser Glu Phe Leu Ile Val Leu Arg Asp Val Val Gly Gly Met Asn									
	110 115 120									
	CAT CTA CGA GAG AAT GGT ATA GTG CAC CGT GAT ATC AAG CCA GGA AAT	492								
	His Leu Arg Glu Asn Gly Ile Val His Arg Asp Ile Lys Pro Gly Asn									

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	125					130					135					140	
				GTT													540
	Ile	Met	Arg	Val		Gly	Glu	Asp	Gly		Ser	Val	Tyr	Lys		Thr	
5					145					150					155		
J	GAT	ттт	GGT	GCA	GCT	AGA	GAA	TTA	GAA	GAT	GAT	GAG	CAG	ттт	GTT	TCT	588
	_			Ala													
	-			160					165					170			
10	CTG	TAT	GGC	ACA	GAA	GAA	TAT	TTG	CAC	CCT	GAT	ATG	TAT	GAG	AGA	GCA	636
	Leu	Tyr	_	Thr	Glu	Glu	Tyr		His	Pro	Asp	Met		Glu	Arg	Ala	
			175					180					185				
	CTC	СТА	ΔCΔ	AAA	СУТ	СУТ	CAG	DAG	444	ጥልጥ	CCA	GCA	ACA	СФФ	GAT	СФФ	684
15				Lys	-												
		190		•	_		195	-	-	•	-	200			_		
	TGG	AGC	ATT	GGG	GTA	ACA	TTT	TAC	CAT	GCA	GCT	ACT	GGA	TCA	CTG	CCA	732
	Trp	Ser	Ile	Gly	Val	Thr	Phe	Tyr	His	Ala		Thr	Gly	Ser	Leu		
20	205					210					215					220	
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	ATA	ATT	ACA	GGA	AAG	CCT	TCT	GGT	GCA	ATA	TCT	GGA	GTA	CAG	AAA	GCA	828
	Ile	Ile	Thr	Gly	Lys	Pro	Ser	Gly	Ala	Ile	Ser	Gly	Val	Gln	Lys	Ala	
				240					245					250			
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	CTT	TCT	CGG	GGT	CTT	CAG	GTT	CTA	CTT	ACC	CCT	GTT	CTT	GCA	AAC	ATC	924
35	Leu	Ser	Arg	Gly	Leu	Gln	Val	Leu	Leu	Thr	Pro	Val	Leu	Ala	Asn	Ile	
		270					275					280					
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	GAA	ACT	AGT	GAT	ATA	CTT	CAC	CGA	ATG	GTA	ATT	CAT	GTT	TTT	TCG	CTA	1020
	Glu	Thr	Ser	Asp	Ile	Leu	His	Arg	Met	Val	Ile	His	Val	Phe	Ser	Leu	
					305					310					315		

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	CAA	CAÃ	ATG	ACA	GCT	CAT	AAG	TTA	TAT	ATA	CAT	AGC	TAT	AAT	ACT	GCT	1068
	Gln	Gln	Met	Thr	Ala	His	Lys	Ile	Tyr	Ile	His	Ser	Tyr	Asn	Thr	Ala	
				320					325					330			
							GTA										1116
5	Thr	Ile		His	Glu	Leu	Val		Lys	Gln	Thr	Lys		Ile	Ser	Ser	
			335					340					345				
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10	ASII	350	GIU	пеп	116	ıyı	355	GIY	nrg	Arg	Бей	360	Dea	014		07	
10		330					333										
	AGG	CTG	GCA	CAA	CAT	TTC	CCT	AAA	ACT	ACT	GAG	GAA	AAC	CCT	ATA	TTT	1212
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	365					370					375					380	
15																	
																AAA	1260
	Val	Val	Ser	Arg	Glu	Pro	Leu	Asn	Thr	Ile	Gly	Leu	Ile	Tyr		Lys	
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				400					405					410			
	AGC	ልጥር	GCT	AAG	GCA	ልጥል	ACA	GGG	GTT	GTG	TGT	ТАТ	GCC	TGC	AGA	ATT	1356
25																Ile	
			415					420			_	_	425	-			
	GCC	AGT	ACC	TTA	CTG	CTT	TAT	CAG	GAA	TTA	ATG	CGA	AAG	GGG	ATA	CGA	1404
	Ala	Ser	Thr	Leu	Leu	Leu	Tyr	Gln	Glu	Leu	Met	Arg	Lys	Gly	Ile	Arg	
30		430					435					440					
																AAA -	1452
			Ile	Glu	Leu			Asp	Asp	Tyr			Thr	Val	Hls	Lys	
25	445	1				450					455					460	
35	220		C 3 3	- CIDIT	o ma	3 000	י אריא	· mmc	CAM	י שישיר	י חורים	ነ አጥሮ	י אכיא	7 A C	ייייר ע	GAA	1500
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40	AAA	ACI	GTO	AAA	GTA	rat .	GAA	AAG	TTG	ATG	AAC	ATC	: AAC	CTG	GAA	GCG	1548
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	_			480					485					490			
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	GC	GAC	TT	A GGT	GAZ	TA A	TCA	GAC	ATA	CAC	: ACC	: AAA	TTC	TTO	AGA	A CTT	1596

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	Ala	Glū	Leu 495	Gly	Glu	Ile	Ser	Asp 500	Ile	His	Thr	Lys	Leu 505	Leu	Arg	Leu	
	TCC	AGT	TCT	CAG	GGA	ACA	ATA	GAA	ACC	AGT	CTT	CAG	GAT	ATC	GAC	AGC	1644
	Ser	Ser	Ser	Gln	Gly	Thr	Ile	Glu	Thr	Ser	Leu	Gln	Asp	Ile	Asp	Ser	
5		510					515					520					
	AGA	TTA	TCT	CCA	GGT	GGA	TCA	CTG	GCA	GAC	GCA	TGG	GCA	CAT	CAA	GAA	1692
	Arg	Leu	Ser	Pro	Gly	Gly	Ser	Leu	Ala	Asp	Ala	$\operatorname{\mathtt{Trp}}$	Ala	His	Gln	Glu	
	525					530					535					540	
10																	
					AAA												1740
	Gly	Thr	His	Pro	Lys	Asp	Arg	Asn	Val		Lys	Leu	Gln	Val		Leu	
					545					550					555		
15	AAT	TGC	ATG	ACA	GAG	ATT	TAC	TAT	CAG	TTC	AAA	AAA	GAC	AAA	GCA	GAA	1788
	Asn	Cys	Met	Thr	Glu	Ile	Tyr	Tyr	Gln	Phe	Lys	Lys	Asp	Lys	Ala	Glu	
				560					565					570			
	CGT	AGA	TTA	GCT	TAT	AAT	GAA	GAA	CAA	ATC	CAC	AAA	TTT	GAT	AAG	CAA	1836
20	Arg	Arg	Leu	Ala	Tyr	Asn	Glu	Glu	Gln	Ile	His	Lys	Phe	Asp	Lys	Gln	
			575					580					585				
	AAA	CTG	TAT	TAC	CAT	GCC	ACA	AAA	GCT	ATG	ACG	CAC	TTT	ACA	GAT	GAA	1884
	Lys	Leu	Tyr	Tyr	His	Ala	Thr	Lys	Ala	Met	Thr	His	Phe	Thr	Asp	Glu	
25		590					595					600					
	TGT	GTT	AAA	AAG	TAT	GAG	GCA	TTT	TTG	AAT	AAG	TCA	GAA	GAA	TGG	ATA	1932
	Cys	Val	Lys	Lys	Tyr	Glu	Ala	Phe	Leu	Asn	Lys	Ser	Glu	Glu	Trp	Ile	
	605					610					615					620	
30																	
	AGA	AAG	ATG	CTT	CAT	CTT	AGG	AAA	CAG	TTA	TTA	TCG	CTG	ACT	AAT	CAG	1980
	Arg	Lys	Met	Leu	His	Leu	Arg	Lys	Gln		Leu	Ser	Leu	Thr		Gln	
					625					630					635		
35	TGT	TTT	GAT	ATT	GAA	GAA	GAA	GTA	TCA	AAA	TAT	CAA	GAA	TAT	ACT	AAT	2028
	Суѕ	Phe	Asp	Ile	Glu	Glu	Glu	Val	Ser	Lys	Tyr	G1n	Glu	Tyr	Thr	Asn	
				640					645					650			
	GAG	TTA	CAA	GAA	ACT	CTG	ССТ	CAG	AAA	ATG	TTT	ACA	GCT	TCC	AGT	GGA	. 2076
40																Gly	
			655					660	_				665			_	
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	670	675	680
5	GAA ATG ACT CTT GGT ATG Glu Met Thr Leu Gly Met 685	Lys Lys Leu Lys Glu	
3	GTT AAA GAA CTT GCT GAA Val Lys Glu Leu Ala Glu 705		
10	TTA ACC ATG GAT GGT GGC Leu Thr Met Asp Gly Gly 720		
15	ATAGAAGTTT AAGAAAAGTT	TCCGTTTGCA CAAGAAAATA	ACGCTTGGGC ATTAAATGAA 2329
	TGCCTTTATA GATAGTCACT	TGTTTCTACA ATTCAGTATT	TGATGTGGTC GTGTAAATAT 2389
	GTACAATATT GTAAATACAT	АААААТАТА CAAATTTTTG	GCTGCTGTGA AAATGTAATT 2449
20	TTATCTTTTA ACATTTATAA	TTATATGAGG AAATTTGACC	TCAGTGATCA CGAGAAGAAA 2509
	GCCATGACCG ACCAATATGT	TGACATACTG ATCCTCTACT	CTGAGTGGGG CTAAATAAGT 2569
25	TATTTCTCT GACCGCCTAC	TGGAAATATT TTTAAGTGGA	ACCAAAATAG GCATCCTTAC 2629
23	AAATCAGGAA GACTGACTTG	ACACGTTTGT AAATGGTAGA	ACGGTGGCTA CTGTGAGTGG 2689
	GGAGCAGAAC CGCACCACTG	TTATACTGGG ATAACAATTT	TTTTGAGAAG GATAAAGTGG 2749
30	CATTATTTTA TTTTACAAGG	TGCCCAGATC CCAGTTATCC	TTGTATCCAT GTAATTTCAG 2809
	ATGAATTATT AAGCAAACAT	TTTAAAGTGA ATTCATTATT	AAAAACTATT CATTTTTTC 2869
35	CTTTGGCCAT AAATGTGTAA	TTGTCATTAA AATTCTAAGG	F TCATTTCAAC TGTTTTAAGC 2929
JJ	TGTATTTCTT TAATTCTGCT	TACTATTTCA TGGAAAAAA	A TAAATTTCTC AATTTTAAAA 2989
	ААААА		2994

(2) INFORMATION FOR SEQ ID NO:2:

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(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 729 amino acids

(B) TYPE: amino acid(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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Met Gln Ser Thr Ser Asn His Leu Trp Leu Leu Ser Asp Ile Leu Gly

1 5 10 15

10 Gln Gly Ala Thr Ala Asn Val Phe Arg Gly Arg His Lys Lys Thr Gly
20 25 30

Asp Leu Phe Ala Ile Lys Val Phe Asn Asn Ile Ser Phe Leu Arg Pro 35 40 45

Val Asp Val Gln Met Arg Glu Phe Glu Val Leu Lys Lys Leu Asn His

Lys Asn Ile Val Lys Leu Phe Ala Ile Glu Glu Glu Thr Thr Thr Arg

20 65 70 75 80

His Lys Val Leu Ile Met Glu Phe Cys Pro Cys Gly Ser Leu Tyr Thr 85 90 95

Val Leu Glu Glu Pro Ser Asn Ala Tyr Gly Leu Pro Glu Ser Glu Phe
100 105 110

Leu Ile Val Leu Arg Asp Val Val Gly Gly Met Asn His Leu Arg Glu 115 120 125

Asn Gly Ile Val His Arg Asp Ile Lys Pro Gly Asn Ile Met Arg Val

Ile Gly Glu Asp Gly Gln Ser Val Tyr Lys Leu Thr Asp Phe Gly Ala 35 145 150 155 160

Ala Arg Glu Leu Glu Asp Asp Glu Gln Phe Val Ser Leu Tyr Gly Thr
165 170 175

Glu Glu Tyr Leu His Pro Asp Met Tyr Glu Arg Ala Val Leu Arg Lys
180 185 190

Asp His Gln Lys Lys Tyr Gly Ala Thr Val Asp Leu Trp Ser Ile Gly
195 200 205

WO 98/39410 PCT/US98/04496 Val Thr Phe Tyr His Ala Ala Thr Gly Ser Leu Pro Phe Arg Pro Phe Glu Gly Pro Arg Arg Asn Lys Glu Val Met Tyr Lys Ile Ile Thr Gly Lys Pro Ser Gly Ala Ile Ser Gly Val Gln Lys Ala Glu Asn Gly Pro Ile Asp Trp Ser Gly Asp Met Pro Val Ser Cys Ser Leu Ser Arg Gly Leu Gln Val Leu Leu Thr Pro Val Leu Ala Asn Ile Leu Glu Ala Asp Gln Glu Lys Cys Trp Gly Phe Asp Gln Phe Phe Ala Glu Thr Ser Asp Ile Leu His Arg Met Val Ile His Val Phe Ser Leu Gln Gln Met Thr Ala His Lys Ile Tyr Ile His Ser Tyr Asn Thr Ala Thr Ile Phe His Glu Leu Val Tyr Lys Gln Thr Lys Ile Ile Ser Ser Asn Gln Glu Leu Ile Tyr Glu Gly Arg Arg Leu Val Leu Glu Pro Gly Arg Leu Ala Gln His Phe Pro Lys Thr Thr Glu Glu Asn Pro Ile Phe Val Val Ser Arg Glu Pro Leu Asn Thr Ile Gly Leu Ile Tyr Glu Lys Ile Ser Leu Pro Lys Val His Pro Arg Tyr Asp Leu Asp Gly Asp Ala Ser Met Ala Lys Ala Ile Thr Gly Val Val Cys Tyr Ala Cys Arg Ile Ala Ser Thr Leu Leu Leu Tyr Gln Glu Leu Met Arg Lys Gly Ile Arg Trp Leu Ile Glu

WO 98/39410 PCT/US98/04496 Leu Ile Lys Asp Asp Tyr Asn Glu Thr Val His Lys Lys Thr Glu Val Val Ile Thr Leu Asp Phe Cys Ile Arg Asn Ile Glu Lys Thr Val Lys Val Tyr Glu Lys Leu Met Lys Ile Asn Leu Glu Ala Ala Glu Leu Gly Glu Ile Ser Asp Ile His Thr Lys Leu Leu Arg Leu Ser Ser Ser Gln . 500 Gly Thr Ile Glu Thr Ser Leu Gln Asp Ile Asp Ser Arg Leu Ser Pro Gly Gly Ser Leu Ala Asp Ala Trp Ala His Gln Glu Gly Thr His Pro Lys Asp Arg Asn Val Glu Lys Leu Gln Val Leu Leu Asn Cys Met Thr Glu Ile Tyr Tyr Gln Phe Lys Lys Asp Lys Ala Glu Arg Arg Leu Ala Tyr Asn Glu Glu Gln Ile His Lys Phe Asp Lys Gln Lys Leu Tyr Tyr His Ala Thr Lys Ala Met Thr His Phe Thr Asp Glu Cys Val Lys Lys Tyr Glu Ala Phe Leu Asn Lys Ser Glu Glu Trp Ile Arg Lys Met Leu His Leu Arg Lys Gln Leu Leu Ser Leu Thr Asn Gln Cys Phe Asp Ile Glu Glu Glu Val Ser Lys Tyr Gln Glu Tyr Thr Asn Glu Leu Gln Glu Thr Leu Pro Gln Lys Met Phe Thr Ala Ser Ser Gly Ile Lys His Thr Met Thr Pro Ile Tyr Pro Ser Ser Asn Thr Leu Val Glu Met Thr Leu

Gly Met Lys Lys Leu Lys Glu Glu Met Glu Gly Val Val Lys Glu Leu 690 695 700

Ala Glu Asn Asn His Ile Leu Glu Arg Phe Gly Ser Leu Thr Met Asp 705 710 715 720

5 Gly Gly Leu Arg Asn Val Asp Cys Leu

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WHAT IS CLAIMED IS:

1. An isolated T2K protein comprising SEQ ID NO: 2 or a fragment thereof having T2K-specific activity.

- 2. An isolated protein according to claim 1, wherein said protein specifically phosphorylates IκB at serine 36.
 - 3. An isolated protein according to claim 1, wherein said protein comprises a deletion mutant of SEQ ID NO:2, said deletion mutant comprising SEQ ID NO:2, residues 1-250 or 251-729.

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- 4. A recombinant nucleic acid encoding a protein according to claim 1.
- 5. A cell comprising a nucleic acid according to claim 4.
- 6. A method of making an isolated T2K protein, comprising steps: introducing a nucleic acid according to claim 4 into a host cell or cellular extract, incubating said host cell or extract under conditions whereby said nucleic acid is expressed as a transcript and said transcript is expressed as a translation product comprising said protein, and isolating said translation product.
- 20 7. An isolated T2K protein made by the method of claim 6.
 - 8. An isolated T2K nucleic acid comprising SEQ ID NO: 1, or a fragment thereof having at least 24 consecutive bases of SEQ ID NO: 1 and sufficient to specifically hybridize with a nucleic acid having the sequence of SEQ ID NO: 1.

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9. A method of screening for an agent which modulates the binding of a T2K protein to a binding target, said method comprising the steps of:

incubating a mixture comprising:

an isolated protein according to claim 1,

a binding target of said protein, and

a candidate agent;

under conditions whereby, but for the presence of said agent, said protein specifically binds said binding target at a reference affinity;

detecting the binding affinity of said protein to said binding target to determine an agentbiased affinity,

wherein a difference between the agent-biased affinity and the reference affinity indicates that said agent modulates the binding of said protein to said binding target.

- 10. A method according to claim 9, wherein said binding target is a substrate comprising IkB serine 36 and said binding affinity is detected as phosphorylation of said IkB serine 36.
- 10 11. A method of screening for an agent which modulates IkB phosphorylation by an IkB kinase specific for IkB serine 36, said method comprising the steps of:

incubating a mixture comprising:

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an isolated IkB serine 36 specific kinase,

a substrate comprising IkB serine 36, and

a candidate agent;

under conditions whereby, but for the presence of said agent, said kinase specifically phosphorylates said substrate at IkB serine 36 at a reference activity;

detecting the phosphorylation of said substrate by said kinase to determine an agentbiased activity,

- wherein a difference between the agent-biased activity and the reference activity indicates that said agent modulates modulates IkB serine 36 phosphorylation.
- 12. A method according to claim 11, wherein said kinase comprises the sequence of KIAA0151 or SEQ ID NO: 2, or a fragment of either which specifically phosphorylates IkB at serine 36.

INTERNATIONAL SEARCH REPORT

International application No. PCT/US98/04496

IPC(6) : US CL :	SSIFICATION OF SUBJECT MATTER C12N 1/14, 1/20, 5/00, 15/00, 9/12; C07H 21/04; C1: 435/15, 194, 252.3, 254.11, 320.1, 325; 536/23.2; 93: o International Patent Classification (IPC) or to both n	/22	cation and IPC					
	DS SEARCHED	· <u></u>						
Minimum do	ocumentation searched (classification system followed	by classificati	on symbols)					
U.S. : 4	435/15, 194, 252.3, 254.11, 320.1, 325; 536/23.2; 935	222						
Documentati	ion searched other than minimum documentation to the	extent that suc	documents are included	in the fields searched				
Electronic d	ata base consulted during the international search (na	ne of data has	and where practicable	search terms used)				
APS: (US	SPAT, JPO and EPOABS); search terms: TRAF and registry, CAPLUS, Medline); search terms: same as ab	ΓRAF2, kinase		, source to the same to the sa				
C. DOC	UMENTS CONSIDERED TO BE RELEVANT							
Category*	Citation of document, with indication, where app	ropriate, of the	relevant passages	Relevant to claim No.				
A	MALININ et al. MAP3K-related kinase involved in NF-kB induction by TNF, CD95 and IL-1. Nature. February 06, 1997. Vol. 385. pages 540-544.							
A,P	SONG et al. Tumor necrosis factor (TNF)-mediated kinase cascades: Bifurcation of nuclear factor-kB and c-jun N-terminal kinase (JNK/SAPK) pathways at TNF receptor-associated factor 2. Proceedings of the National Academy of Sciences USA September 1997. Vol. 94. pages 9792-9796.							
A,P	NATOLI et al. Tumor necrosis facto downstream of TNF receptor-associate Vol. 272. No. 42. pages 26079-2608	1-12						
Furtl	her documents are listed in the continuation of Box C	Se	e patent family annex.					
"A" do	pecial categories of cited documents: Decument defining the general state of the art which is not considered be of particular relevance	date		ternational filing date or priority blication but cited to understand e invention				
L do	wher document published on or after the international filing date occument which may throw doubts on priority claim(s) or which is ted to establish the publication date of another citation or other	consi		ne claimed invention cannot be ered to involve an inventive step				
O do	the claimed invention cannot be e step when the document is ch documents, such combination the art							
	ocument published prior to the international filing date but later than te priority date claimed	*&* docu	ment member of the same pate	nt family				
Date of the	e actual completion of the international search	Date of maili	ng of the international se	' '				
Commissi Box PCT	mailing address of the ISA/US oner of Patents and Trademarks on, D.C. 20231 No. (703) 305-3230	Authorized o EINAR S Telephoneur	TOLE / / / / / / / / / / / / / / / / / / /	ollent				
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